

## TEST REQUEST FORM

Name \_\_\_\_\_

Affiliation \_\_\_\_\_

Address \_\_\_\_\_

Tel. \_\_\_\_\_

Fax \_\_\_\_\_

e-mail \_\_\_\_\_

### Requested test

- |  |   |
|--|---|
| <input type="checkbox"/> Detection of anti- <i>Trichinella</i> sp. antibodies in swine sera by indirect ELISA (test method MI-01)  | N° samples _____                          |
| <input type="checkbox"/> Detection of anti- <i>Trichinella</i> sp. antibodies in human sera by indirect ELISA (test method MI-03)  | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Trichinella</i> sp. larvae at the species level by Multiplex PCR (test method MI-02)   | N° samples _____                          |
| <input type="checkbox"/> Detection of <i>Trichinella</i> larvae in muscle tissues Reg. EU 2015/1375 10/08/2015, Ann. 1. chap. 1 and Ann. 3   | N° samples _____                          |
| <input type="checkbox"/> Identification of Anisakidae larvae at the species level by PCR/RFLP (test method MI-04)  | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Cryptosporidium</i> oocysts at the species level by PCR/RFLP (test method MI-06)   | N° samples _____                          |
| <input type="checkbox"/> Detection of anti- <i>Opisthorchis</i> antibodies in human sera (test method MI-07)   | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Opisthorchis</i> spp eggs by PCR (test method MI-08)   | N° samples _____                          |
| <input type="checkbox"/> Identification at assemblage level of <i>Giardia duodenalis</i> cysts by PCR/RFLP (test method MI-09)   | N° samples _____                          |
| <input type="checkbox"/> Identification of Anisakidae larvae at species level by Multiplex PCR (test method MI-10)   | N° samples _____                          |
| <input type="checkbox"/> Identification of assemblage A and B of <i>Giardia duodenalis</i> by PCR (test method MI-11)  | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Toxoplasma gondii</i> DNA in food matrices (fresh or processed meat) by LAMP (test method MI-12)   | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Trichinella</i> spp. proteins recognized by specific IgG from serum samples of infected swine by Western blotting (test method MI-13; confirmatory method to be applied on samples tested positive for MI-01 method) | N° samples only positive for MI-01 method |
| <input type="checkbox"/> Identification of <i>Toxoplasma gondii</i> DNA in vegetables by LAMP (test method MI-14)  | N° samples _____                          |
| <input type="checkbox"/> Identification of <i>Echinococcus granulosus sensu lato</i> complex at species/genotype level by PCR/RFLP and Multiplex PCR (test method MI-15)   | N° samples _____                          |
| Sample/s: <input type="checkbox"/> frozen, <input type="checkbox"/> ethyl alcohol, <input type="checkbox"/> alcohol removed before sending   |   |
| <input type="checkbox"/> Other test* (specify) _____   | N° samples _____                          |

**NB: methods marked with \* are not accredited by ACCREDIA**

Client sampling on	
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The test report should be sent by:     post     fax     e-mail     other (specify)

Notes

NOTES

### TEST REQUEST FORM

1. Samples should be sent to: **EURL for Parasites (attention Dr. S.M. Cacciò) - Istituto Superiore di Sanità, viale Regina Elena 299, 00161 Rome, Italy**
2. Sampling procedures, shipping and packaging conditions
  - a. Sera – Sera, collected from non-haemolysed blood samples, should be sent in plastic vials hermetically sealed, refrigerated if delivered within 24 hours, frozen for longer shipping periods. Each vial should be marked with an identification code and the code list should be attached to the present request. Please, keep in mind that the antibody titre decreases at each freezing/thawing process.
  - b. *Trichinella* sp. larvae – Larvae should be collected from human and animal infected muscles by pepsin-HCl digestion and preserved in  $\geq 90\%$  ethyl alcohol. Each isolate (from 1 to thousand of larvae) should be sent at room temperature in a plastic vial hermetically sealed (we suggest to use 1ml vial with screw plug or a conical vial with pressure plug; in both cases sealed by parafilm). Vials should be entirely filled by ethyl alcohol, to avoid larvae to stick on the vial wall. Each vial should be marked with an identification code and the code list should be attached to the present request. We recommend to mark somehow the code to avoid it to be deleted by a leak of alcohol.
  - c. Muscle tissues – Muscle tissues should be collected according to the Annex I (Chapter 1, section 2) and Annex III of the Regulation EU 2015/1375. Provide no less than 10 gr of striated muscle tissues free of all fat and fascia. If the sample is delivered within 48 hours, it can be sent without refrigeration; for longer period of time, it should be sent refrigerated, under vacuum or in a 0.1% merthiolate solution.
  - d. Anisakidae larvae – Larvae should be collected from human and animal infected muscles and preserved in  $\geq 90\%$  ethyl alcohol. Each isolate (from 1 to thousand of larvae) should be sent at room temperature in a plastic vial hermetically sealed (we suggest to use 1-5 ml vial with screw plug sealed by parafilm). Vials should be entirely filled by ethyl alcohol, to avoid larvae to stick on the vial wall. Each vial should be marked with an identification code and the code list should be attached to the present request. We recommend to mark somehow the code to avoid it to be deleted by a leak of alcohol.
  - e. Cystic material – cystic material should be preserved frozen or in ethyl alcohol ( $\geq 70\%$ ), the alcohol may be removed before sending. Each sample should be sent at room temperature or refrigerated in a plastic vial hermetically sealed (sealing by parafilm). Each vial should be marked with an identification code and the code list should be attached to the present request. We recommend to mark somehow the code to avoid it to be deleted by a leak of alcohol. Cystic material can be hydatid cyst, protoscolexes and germinal layer. The whole hydatid cyst and protoscolexes are considered tissues of choice for the analysis.
  - f. Human and animal faeces – faecal material (minimum 1 mL) has to be preserved by adding an equal volume of  $\geq 90\%$  ethyl alcohol. Each sample should be sent at room temperature in a plastic vial hermetically sealed (we suggest to use 1-5 mL vial with screw plug sealed by parafilm). Each vial should be marked with an identification code and the code list should be attached to the present request. We recommend to mark somehow the code to avoid it to be deleted by a leak of alcohol.
  - g. Fresh or processed meat (not seasoned) – Each 5 gr sample, should be sent refrigerated (temperature  $\leq 14^{\circ}\text{C}$ ) in 50 ml polypropylene conical vial, with screw plug. Each vial should be marked with an identification code and the code list should be attached to the present request.
  - h. Leafy vegetables - Each 50 gr sample, should be sent refrigerated (temperature between  $4^{\circ}\text{C}$  and  $14^{\circ}\text{C}$ ) in sample plastic bag, appropriately sealed. Each bag should be marked with an identification code and the code list should be attached to the present request.
3. Samples should be delivered to the laboratory between 8 am and 5 pm of every working day from Monday to Friday. We suggest to send the samples at the beginning of the week, be careful of possible midweek Italian holidays.
4. Sampling, packaging and shipping of samples to this laboratory are under the responsibility of the applicant. For any further information, please contact us (Tel. +39 06 49902304/2308/2310/3017/3379; Fax: +39 06 49903561; or by email: [simone.caccio@iss.it](mailto:simone.caccio@iss.it)).
5. Maximum delay for test report forward is 60 days from sample receipt, but it may be otherwise agreed.
6. Sample storage period for further analysis is 30 days. After this time, samples are destroyed or returned to the applicant only on request.
7. The original raw data and a copy of the test report are kept for 10 years at this Institute.
8. At present, this service is free of charge, but for sample shipping cost.
9. The accreditation, according to the ISO/IEC 17025 international standard, is submitted to a convention and recognizes the technical competence of the laboratory to perform specific analytical methods. The accreditation body, ACCREDIA ([www.accredia.it](http://www.accredia.it)), does not take any responsibility on the acceptance of the test sample and on the test results or their interpretation.

Date \_\_\_\_\_

Signature of the applicant \_\_\_\_\_

#### **Do not fill in this section**

Sample received on \_\_\_\_\_

Condition of sample at arrival    suitable     unsuitable

Notes